











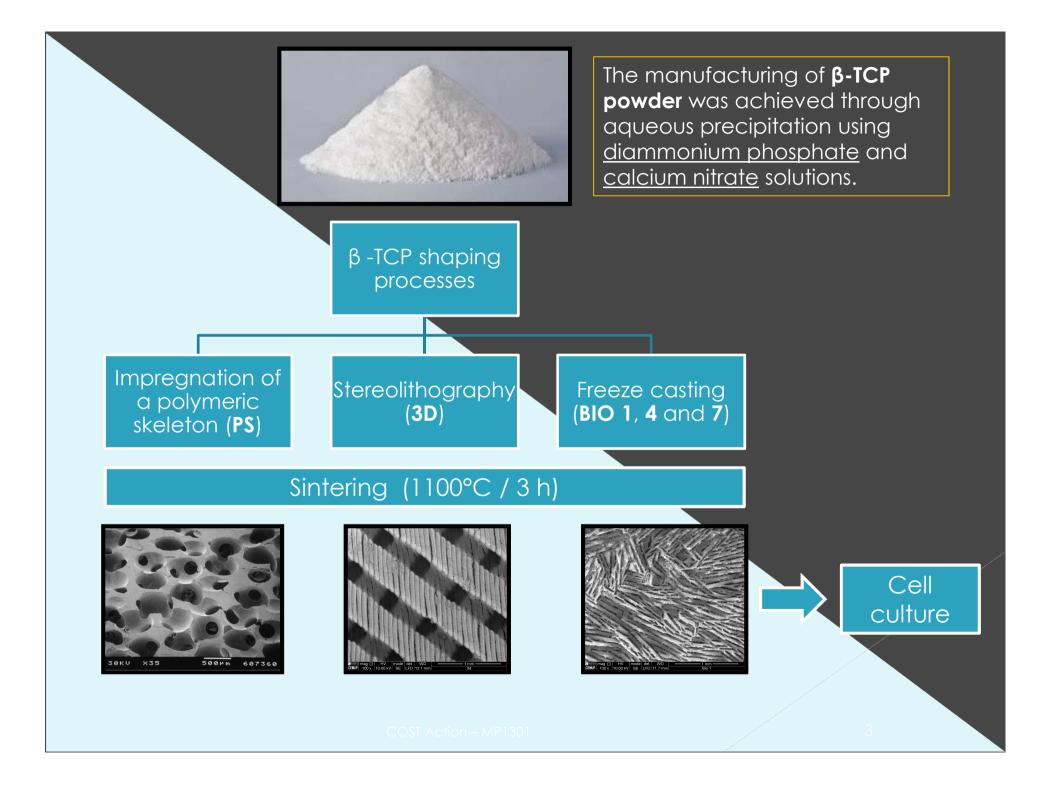
### **β-TCP porous scaffolds : part 2. Relation between** structure and cell invasion

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- This study sets out to explore the links between <u>cell colonization</u> (Osteoblasts and endothelial cells) and the <u>porous architecture</u> of a frozen β-TCP bone implant in vitro.
- Shaping processes were chosen for their ability to generate original porous structures:
  - > Spherical interconnected pore network for polymeric impregnation
  - > <u>Cubical interconnected pore network</u> for stereolithography
  - > Ellipsoidal tubular interconnected pores for freeze casting
- Samples obtained by impregnation of a polymeric skeleton will serve as comparison standard.



#### • Sample characteristics:

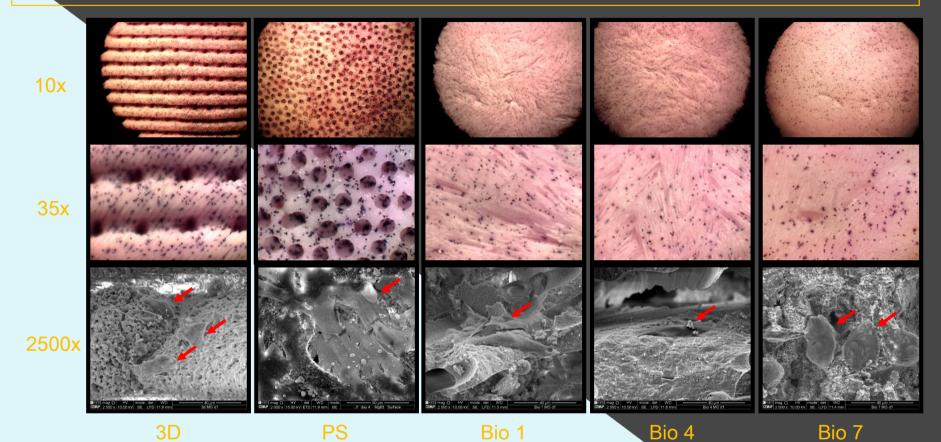
Sample denomination		Shaping method	Porosity (%)	Pore and interconnection diameter (µm)
3D		stereolithography	50	500 / 100
PS		Impregnation of a polymeric skeleton	65	400 – 500 / 100
BIO 1 BIO 4 BIO 7		Freeze casting	50 50 36	150 / 40 360 / 55 150 / 45
Osteoblasts: MG63       2 x 10 <sup>5</sup> cells / well         Endothelial cells: HUVEC       3 mm         COST Action - MP130       3 mm				

All samples were incubated without cells for various periods of time and then seeded with osteoblasts (MG63) in order to determine incubation duration:

Incubation period (h)	Observations		
1	High cell mortality : no cell survival past Day 3		
5	High cell mortality : no cell survival past Day 3		
24	High cell survival: positive response noticed		
SDFSSO </th <th><ul> <li>Final</li> <li></li></ul></th>	<ul> <li>Final</li> <li></li></ul>		

SEM pictures taken at Day 1 and 3 (post seeding), after 24 h of incubation with culture medium. High magnification (x 5000) allowed morphological examination of cells

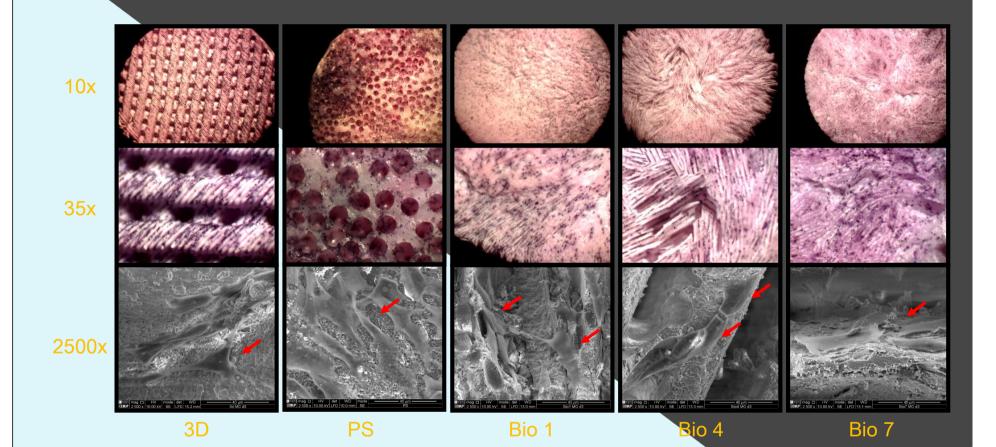
#### M and SEM observations – DAY 1



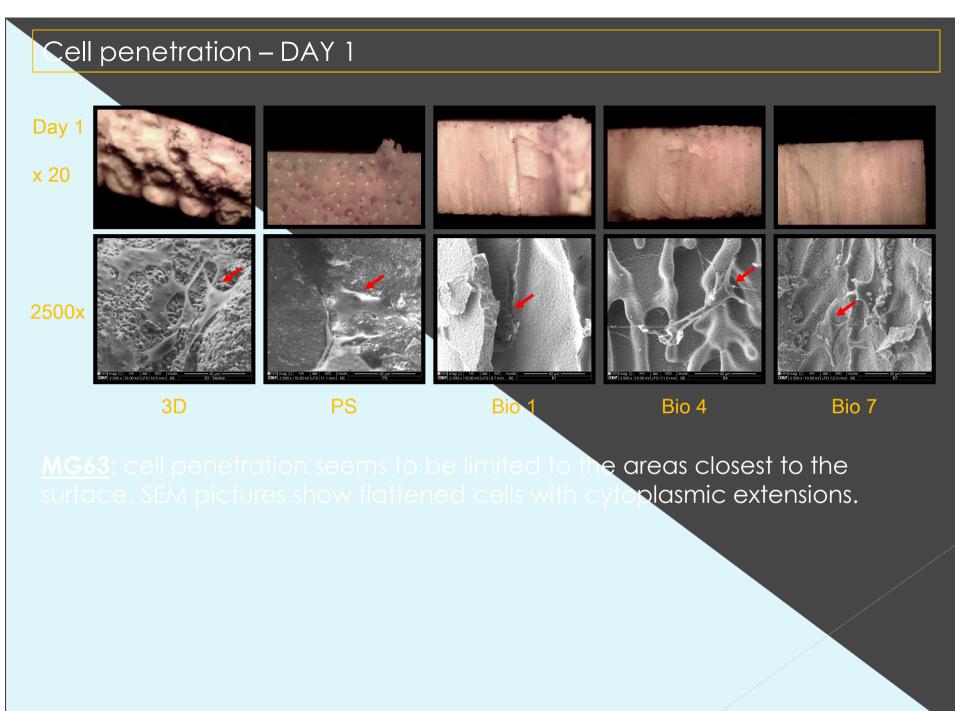
<u>MG63</u>: Osteoblasts responded well. MTT stained cells can be seen all over the samples. Their morphology suggest that they are adherent to the material.

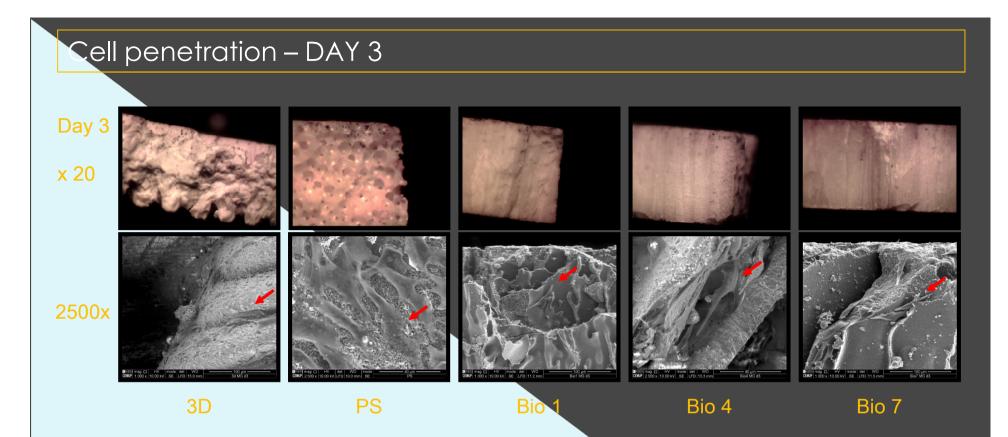
**HUVECs**: Endothelial cells responded poorly. Control well seems to indicate that the material is not responsible for this response. (Results not shown)

#### M and SEM observations – DAY 3



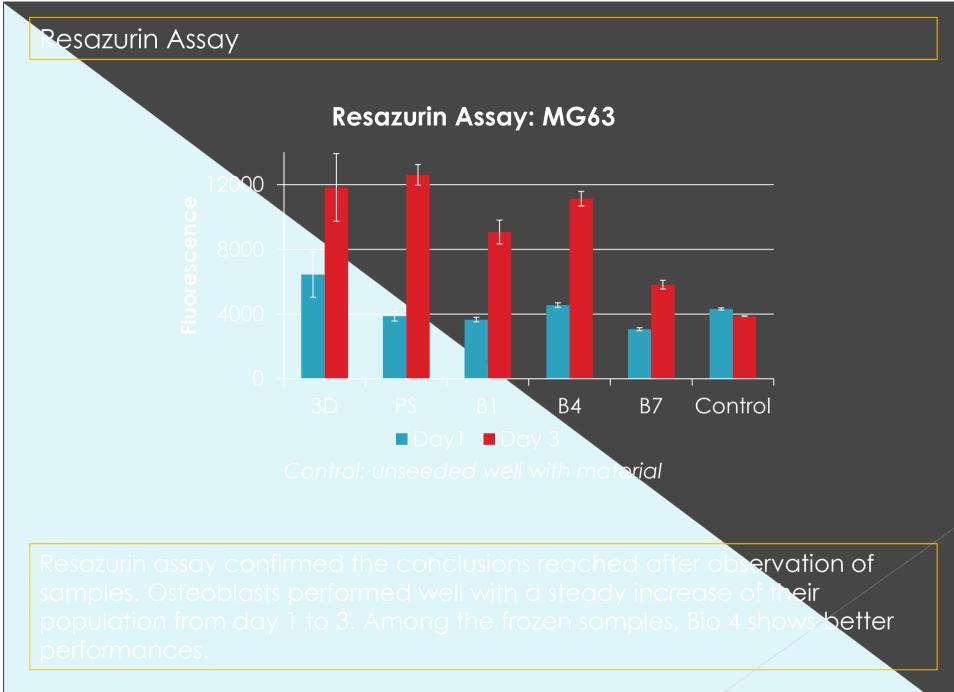
MG63: cell population increased on all the samples. However, 3D and PS samples seem to have a denser cell population. HUVECs: No endothelial cells were observed. (Results not shown)

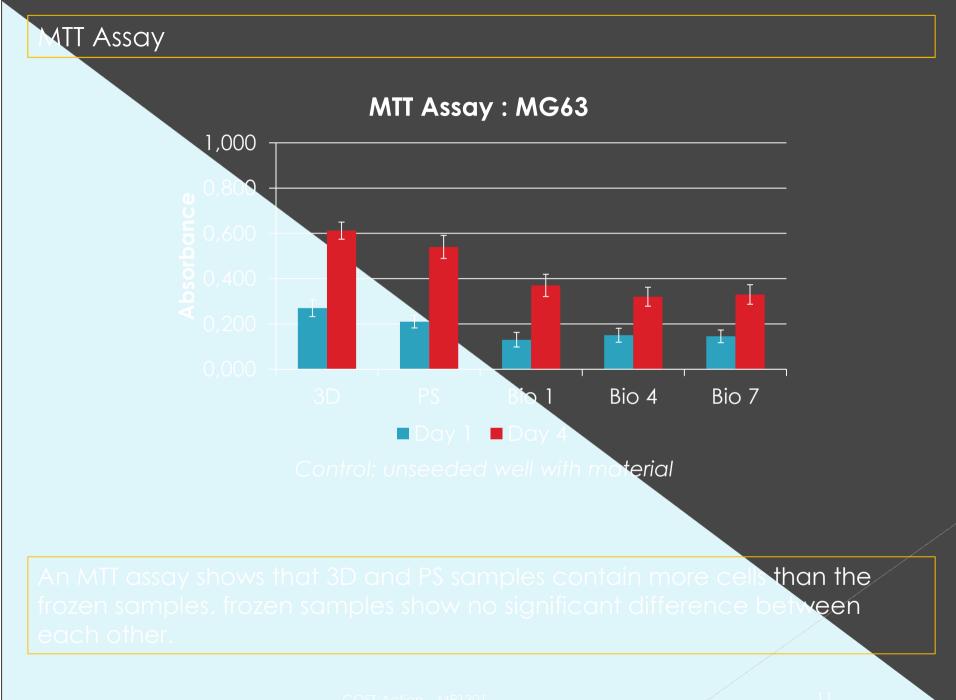




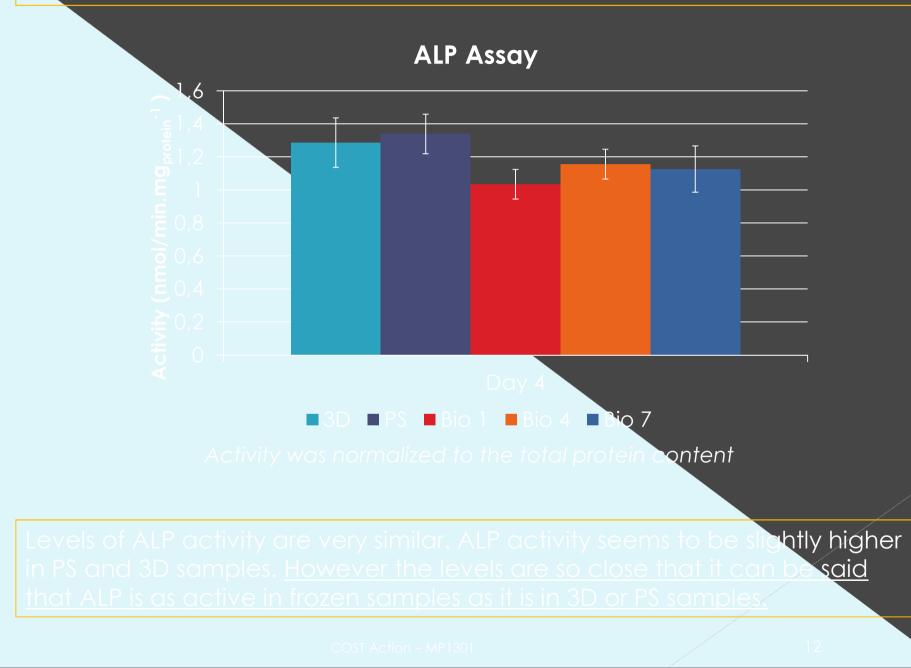
<u>MG63</u>: cell populations found inside the samples were larger than on the first day. Their morphology suggest that they were adherent and well adapted.

Cell penetration in PS and 3D samples is restricted to the pores closest to their surface whereas frozen sample had cell deep into their porous network (Bio 4). Global porosity does not seem to have an impact on cell penetration.





#### Nkaline Phosphatase Assay



## Conclusion

- Shaping techniques allowed the fabrication of samples exhibiting original structures.
- Bioactivity of  $\beta$ -TCP did not seem to be altered by shaping processes
- Osteoblasts reacted well to the material (Flat adherent cells with cytoplasmic extensions)
- 3D and PS samples hosted the largest number of cells at day 4 (MTT Assay).
- Cells penetrated further into the frozen samples (Bio 4), it could be explained by the presence of the local tubular structure and its diameter.
- Frozen samples (Bio 1, 4 and 7) exhibited levels of alkaline phosphatase activity as high as in 3D and PS samples.

## Outlooks

- These preliminary tests and results highlighted several issues and allowed for the fine tuning of the cell culture protocols for future experimentation.
- The next step of this study will be to perform the above tests over longer periods of time (7 to 21 days).
- Future tests: endothelial cell culture, co-cultures, gene activity (RT-PCR), in vivo tests...

# Acknowledgements











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